

STAT-5 and STAT-6 in Breast Cancer: Potential Crosstalk With Estrogen and Progesterone Receptors Can Affect Cell Proliferation and Metastasis

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Abstract

Background: Signal transducers and activators of transcription 5a and 6 (STAT5a and STAT6) play a critical role in tumorigenesis of mammary glands. Based on previous studies, the breast cancer is largely dependent on hormone receptors. Consequently, it is very interesting to decipher the relationship between the STAT5a and STAT6 expression and the molecular distribution of estrogen receptors (ERs) and progesterone receptors (PRs) in mammary tumors.

Methods: Our study analyzed the expression of STAT5a and STAT6, ER α , ER β and PR in 40 breast tumor tissues using quantitative real-time polymerase chain reaction (qRT-PCR). Furthermore, the Ki-67 and HER2 status were detected using immunohistochemistry.

Results: STAT5a and STAT6 were retained in the majority of the cases studied. Increasing of STAT5a and STAT6 is significantly associated with ERs and PR. The coexpression of both STAT5a and STAT6 with ERs and PR is associated with high tumor grades. Moreover, the coexpression of STAT5a and STAT6 with ER α and PR is associated with a high proliferation index. In addition, (STAT6 + ER β +) and (STAT6 + PR+) breast cancer subgroups are associated with lymph node infiltration (P = 0.001 and P = 0.03, respectively).

Conclusions: Our study results provide an interaction between STAT5a and STAT6 with ERs and PR inducing cell proliferation. Coexpression of STAT5a and STAT6 with ERs and PR can predict sensibility to hormonal therapy.

Keywords: Breast cancer; STAT5a; STAT6; ERs; PR; Crosstalk

Introduction

Mammary gland development occurs through the activation of a variety of transcription factors. Inappropriate or constitutive activation of many of these transcription factors is found in breast cancer and may contribute directly to its pathogenesis [1]. Particularly, signal transducers and activators of transcription (STAT), a family of transcription factors, play crucial roles in many cellular functions and are often activated unsuitably in cancer [2]. STATs are latent transcription factors that localized in the cytoplasm. Upon their activation by tyrosine phosphorylation, STATs can dimerize, translocate to the nucleus, bind to DNA, and modulate transcription. Thereby, they regulate cellular functions such as survival, proliferation, and differentiation [3]. Two STAT family members, STAT5 and STAT6, play important roles in normal mammary gland development and both have been implicated in breast tumorigenesis [4].

STAT5, which includes two homologous proteins, STAT5a and STAT5b, plays a crucial role in normal mammary gland development and carcinogenesis. Indeed, STAT5 is activated during post-pubertal mammary gland development, but it was observed to cause epithelial hyper-proliferation and generates precocious functional alveoli formation in virgin mice [5]. Deficient STAT5 mice exhibit a lack of lobulo-alveolar growth of mammary glands and females are unable to lactate [6]. In addition, STAT-5a mediates the prolactin-induced milk protein gene transcription during lactation [7]. The immunohistochemistry study in a large cohort of malignant breast tumors shows that STAT5 was constitutively activated [8]. In addition, Cotarla et al (2004) found that STAT5a is activated in a high proportion of breast cancers [9]. In fact, STAT5a is phosphorylated in nearly 70% of human malignant breast tumors. Furthermore, murine models also support the idea of the role played by STAT5 in mammary tumorigenesis. Mice that express a constitutively activated form of STAT5 develop mammary carcinomas, whereas mice that lack STAT5a are protected against mammary tumors induced by transforming the growth factor α [10]. Taken together, these data imply that

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STAT5 plays a main role in both normal and neoplastic mammary functions.

STAT6 also plays a critical role in mammary gland development and in breast cancer. During early pregnancy, in response to the upregulation of Th2 cytokines (interleukin (IL)-4 and IL-13), STAT6 stimulates alveolar differentiation and proliferation through the induction of the GATA3 expression [11]. STAT6 is instrumental in regulating the balance between Th1 and Th2 cells. It promotes tumor invasiveness and metastasis through the promotion of the Th2 cytokines profile. Moreover, STAT6 can inhibit immunosurveillance against primary solid tumors and metastatic disease. In fact, STAT6-deficient mice are resistant to mammary carcinoma and reject metastatic immortal tumor cells [12].

On the other hand, normal mammary development and breast cancer are largely dependent on other factors such as estrogen receptors (ERs) and progesterone receptors (PRs). In fact, ERs and PRs are critical mediators for mammary gland proliferation after puberty [13]. Furthermore, ERs and PRs may induce cell proliferation and metastasis in breast cancer [14-16]. The present study, therefore, attempts to focus on the assessment of the STAT5a and STAT6 expression levels in malignant breast tumors according to ER and PR distribution.

Materials and Methods

Patients and samples

Malignant mammary tumors were excised during tumorectomies from 40 women (mean age 58.5, range 32 - 85 years) at the Saleh Azaiez Oncology Institute of Tunisia. The women did not undergo any treatment before surgery. The presence of malignant cells in the samples was determined by a pathology specialist. Each diagnosed sample was divided into two portions: one portion was immediately processed for immunohistochemistry and the other portion was frozen and maintained at - 80 °C until RNA extraction. All the procedures followed were examined and approved by the Saleh Azaiez Oncology Institute Ethics Committee. The study was conducted in compliance with the ethical standards of the responsible institution on human subjects as well as with the Helsinki Declaration.

RNA extraction

RNA was extracted from breast specimens by using a mechanical stirrer in the presence of a lysis buffer prior to the use of the total RNA isolation and high pure RNA isolation kits (Roche Diagnostics, Basel, Switzerland) according to the manufacturer's protocol.

cDNA synthesis

The mRNA concentration was measured with a NanoDrop

spectrophotometer, the cDNA synthesis was done in a total volume of 20 µL using the PrimeScript™ first strand cDNA Synthesis Kit (Takara Biotechnology Co., Ltd., Dalian, China) according to the manufacturer's protocol. Mix1 was prepared containing 1 µg of RNA, 2 µL of random hexamer primers, 1 µL of dNTP mixture and RNase-free water (total volume 10 µL). The samples were then incubated for 5 min at 65 °C. Subsequently, Mix2 was prepared containing Mix1, 4 µL of 5 × PrimScript buffer, 0.5 µL of RNase inhibitor, 1 µL of PrimScriptRTase and 4.5 µL of RNase-free water (total volume 20 µL). The samples were incubated at 30 °C for 10 min and at 42 °C for 60 min, and reverse transcriptase were inactivated by heating at 95 °C for 5 min.

PCR amplification

Quantitative real-time polymerase chain reaction (qRT-PCR) was carried out using an ABI Prism 7700 Sequence Detection System (Applied Biosystems; Thermo Fisher Scientific, Inc., Waltham, MA, USA). Specific primers were STAT5a-forward, 5'-CACAGATCAAGCAAGTGGTC-3'; STAT5a-reverse, 5'-CTGTCCATGGTCGGCGTAA-3' and STAT6-forward, 5'-CCTCGTCACCAGTTGCTT-3'; STAT6-reverse, 5'-TCCAGTGCTTTCTGCTCC-3'. The 36B4 expression was used as a control to normalize the data. Primer sequences used to amplify 36B4 were 5'-AATCTCCAGGGGCACCATT-3' and reverse 5'-CGCTGGCTCCCACCTTTGT-3'. Relative mRNA levels were calculated based on the CT values and corrected for the 36b4 expression according to the equation $2^{-\Delta\Delta CT}$ [17]. Relative mRNA levels in the control tissue were equated to 1 and the other values were expressed relative to this. Experiments were performed in triplicate for each data point.

Immunohistochemistry

The immunohistochemistry expression of the oncoprotein Her2/neu and the proliferation index Ki67 were tested on the same set of tumors. The primary antibodies used were: mouse anti-human Her2 (CB11) and mouse anti-human Ki67 (MM1) (NovoCastra, Newcastle, UK). After deparaffinization Sections were subsequently hydrated, incubated for 30 min in 1% hydrogen peroxide to block endogenous activity, and then antigen retrieval was performed by incubating the sections in a 0.01 M citrate buffer (epitope retrieval solution pH 6.0; Leica Microsystems GmbH, Wetzlar, Germany) for 30 min at 98 °C. The primary antibodies were applied for 1 h at 4 °C, with a dilution of 1:40 for Her2 and 1:200 for Ki-67. The sections were then incubated at room temperature with post primary block for 30 min to block nonspecific polymer binding. The sections were incubated with a NovoLink™ Polymer for 30 min at room temperature, followed by incubations with 3,3'-diaminobenzidine (DAB) working solution for 5 min at room temperature to develop peroxidase activity. The slides were counterstained with hematoxylin and mounted. Staining specificity was checked using negative controls. Subsequently, the primary antibodies were applied for 1 h at

Table 1. Relationship Between the mRNA STAT5a and STAT6 Expressions and the Standard Clinical and Pathological Factors and Molecular Settings

Characteristics	Total population	STAT5a		P value	STAT6		P value ^a
		Negative (%)	Positive (%)		Negative (%)	Positive (%)	
Total	40 (100%)	5 (12.5)	35 (87.5)		9 (22.50)	31 (77.50)	
Menopausal status							
Pre-	15 (37.5)	0 (0.0)	15 (42.85)		1 (11.11)	14 (45.16)	
Post-	25 (62.5)	5 (100)	20 (57.14)	NS	8 (88.88)	17 (54.83)	NS
Grade ^b							
SBRI	9 (21.87)	3 (60)	6 (17.14)		5 (55.55)	4 (12.90)	
SBRII	21 (52.50)	0 (0.00)	21 (60.00)	0.0005	4 (44.44)	17 (54.83)	0.002
SBRIII	10 (25)	2 (40)	8 (22.85)		0 (0.0)	10 (32.25)	
Lymph node status							
Positive	21 (52.50)	2 (40)	19 (54.28)		0 (100.0)	21 (67.74)	
Negative	19 (47.50)	3 (60)	16 (45.71)	NS	9 (100)	10 (32.25)	0.01
Tumor size							
≤ 30 mm	27 (67.50)	1 (20)	26 (74.28)	0.0001	4 (44.44)	23 (74.19)	0.0003
> 30 mm	13 (32.50)	4 (80)	9 (25.71)		5 (55.55)	8 (25.80)	
HER2 status							
Positive	10 (25)	0 (00)	10 (28.57)		1 (11.11)	9 (29.03)	
Negative	30 (75)	5 (100)	25 (71.42)	0.0008	8 (88.88)	22 (70.96)	0.002
Ki67							
≤ 14%	15(37.50)	2 (40)	13 (42.85)		2 (22.22)	13 (41.93)	
> 14%	25 (62.50)	3 (60)	22 (57.14)	NS	7 (77.77)	18 (58.06)	NS

^aChi-square test in R. ^bScarff-Bloom-Richardson classification. STAT: signal transducers and activators of transcription; SBR: Scarff-Bloom-Richardson; NS: not significant.

4 °C, with a dilution of 1:40 for Her2 and 1:200 for Ki-67. The sections were then incubated at room temperature with post primary block for 30 min to block nonspecific polymer binding. The sections were incubated with a NovoLink™ Polymer for 30 min at room temperature, followed by incubations with 3,3'-diaminobenzidine (DAB) working solution for 5 min at room temperature to develop peroxidase activity. The slides were counterstained with hematoxylin and mounted. Staining specificity was checked using negative controls. This part has been previously described by Oueslati et al, 2017 [18].

Statistical analysis

The Chi-square test with the R (i386 3.2.1) software was used to compare the quantitative PCR results of STAT5a and STAT6 with clinic-pathological characteristics and with the ERs and PR status. The *t*-test and Mann-Whitney test with GraphPad Prism software were used to compare the STAT5a and STAT6 mRNA level with the clinical and molecular settings. Data are presented as the mean ± standard deviation. P < 0.05 was considered to indicate a statistically significant difference.

Results

STAT5a and STAT6 mRNA expressions and their relationship with clinical and molecular settings

STAT5 and STAT6 have both been reported to be activated in breast tumors and also to have distinct roles in the normal mammary gland and in cancerogenesis. On the other hand, the normal and the pathological mammary glands strongly depend on hormonal receptors. Consequently, in this paper the STAT5a and STAT6 mRNA expressions were evaluated using the qRT-PCR technique in a cohort of malign breast tumors as well as their relationship with ERs and PRs. Our results show a high positivity of both STAT5a and STAT6 (Table 1). The STAT5a and STAT6 expressions were not significantly linked to the menopausal status. The analysis of the mRNA expression level of STAT5a and STAT6 revealed a significant increase of STAT5a in postmenopausal breast tumors (P = 0.03) (Fig. 1). On the contrary, the STAT6 mRNA expression level was three times higher in premenopausal breast tumors compared to postmenopausal breast tumors (P = 0.02) (Fig. 1). When compared to the tumor grade, there was a significant

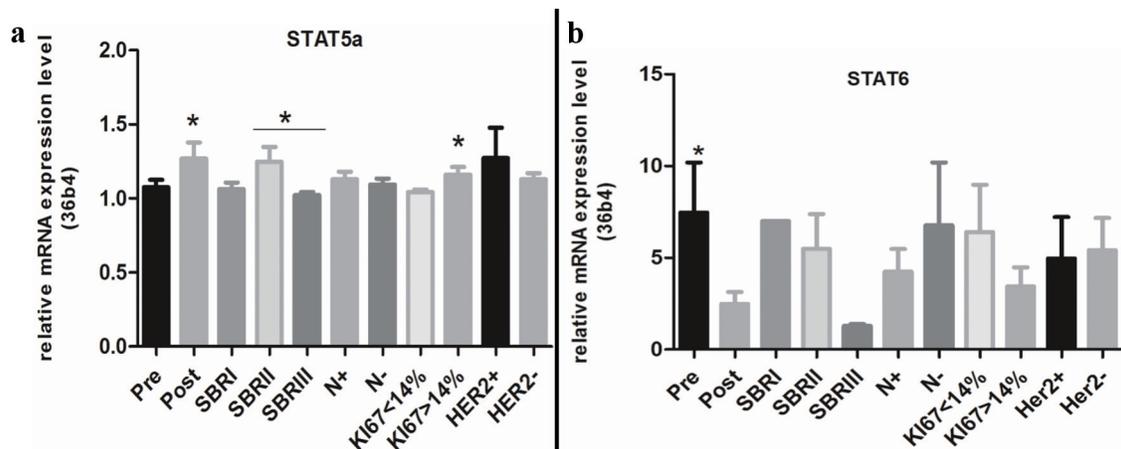


Figure 1. Relative mRNA expression levels of STAT5a and STAT6 according to clinical and molecular information. STAT: signal transducers and activators of transcription; SBR: Scarff-Bloom-Richardson.

association between the STAT5a expression and the Scarff-Bloom-Richardson (SBR)II tumor grade ($P = 0.0005$) (Table 1); while STAT6 is associated with a high tumor grade (SBRII and SBRIII) ($P = 0.002$) (Table 1).

The analysis of the STAT5a mRNA level shows a significant difference between the SBRII and SBRIII tumor grades ($P = 0.01$) (Fig. 1). Our results summarized in Table 2 also show a significant association of the STAT6 (not STAT5a) expression with node infiltration (metastasis) ($P = 0.01$).

Both STAT5a and STAT6 were correlated with a tumor size smaller than 30 mm ($P = 0.0005$ and $P = 0.0001$, respectively). A significant negative association of the STAT5a and STAT6 overexpressions with the HER2/neu status ($P = 0.003$ and $P = 0.004$, respectively) was also found but not with the mRNA levels of STAT5a and STAT6. For Ki-67 (Fig. 2) there is not a significant association of the STAT5a and STAT6 expressions with the proliferation index; however, our results show an association between the increase of the STAT5a mRNA level (but not for STAT6) with a Ki-67 index $> 14\%$ ($P = 0.04$) (Fig. 1).

Relationship between the STAT5a and STAT6 expressions and the ER α , ER β and PR status

In order to determine the relationship between the STAT5a and STAT6 expressions and hormonal receptors, the coex-

Table 2. Relationship Between the STAT5a, STAT6 and ER α , ER β and PR Expressions in Mammary Malign Tumors

	STAT5a+ (n = 35)	P value	STAT6+ (n = 31)	P value
ER α +	24/35	0.004	21/31	0.01
ER β +	23/35	0.01	20/31	0.04
PR+	27/35	0.0000168	25/31	0.00000483

STAT: signal transducers and activators of transcription; ER: estrogen receptor; PR: progesterone receptor.

pression was analyzed (Table 2). The STAT5a and STAT6 expressions are significantly associated ($P = 0.01$ and $P = 0.02$, respectively) with ER α + (but not with ER β +). Furthermore,

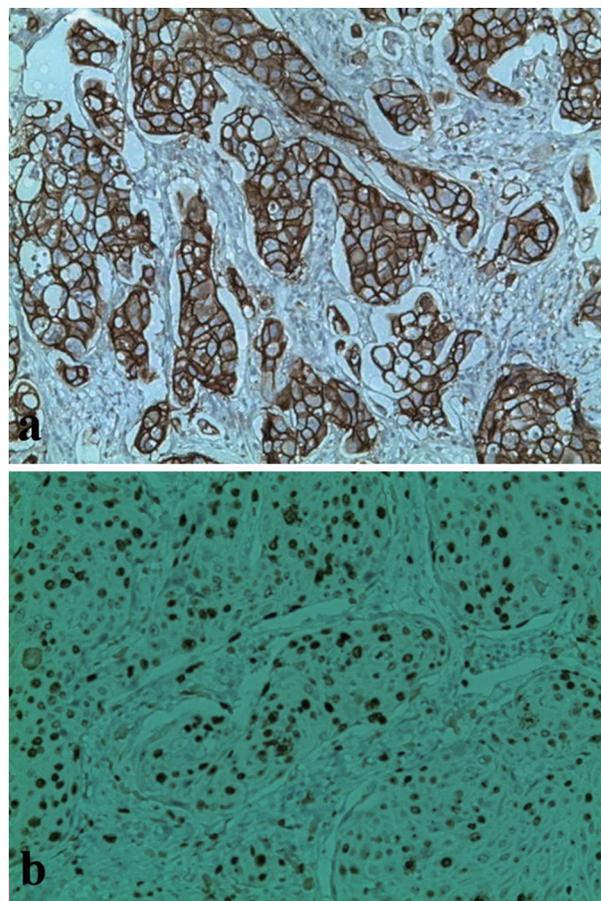


Figure 2. Immunohistochemical staining with anti-HER2 and anti-Ki-67 antibodies in breast tumors. (a) Presence of overexpression of HER2 oncoprotein (HER2 score: 3; original magnification, $\times 400$). (b) Ki-67 proliferation index estimated at 70% (original magnification, $\times 250$).

Table 3. Coexpression of STAT5a With ER α , ER β and PR and the Relationship With Clinical and Molecular Settings

	STAT5a+					
	ER α + (n = 24)	P value	ER β + (n = 23)	P value	PR+ (n = 27)	P value
Menopausal status						
Pre-	10		9		11	
Post-	14	NS	14	NS	16	NS
SBR grade						
I	5	0.0001	4	0.0000365	5	0.0000133
II + III	19		19		22	
Lymph node status						
N+	13	NS	16	0.01	16	NS
N-	11		7		11	
KI67						
> 14	16	0.04	15	NS	19	0.006
\leq 14	8		9		8	
HER2						
+	7		5		6	
-	17	0.009	18	0.0004	21	0.0001

STAT: signal transducers and activators of transcription; ER: estrogen receptor; PR: progesterone receptor; SBR: Scarff-Bloom-Richardson; NS: not significant.

the STAT5a and STAT6 expressions are strongly associated with PR+ (P = 0.000061 and P = 0.00000606, respectively) (Table 2).

Relationship between (STAT5a + ER α +), (STAT5a + ER β +) and (STAT5a + PR+) breast cancer subgroups and clinical settings

Our results (Table 3) reveal the absence of a significant association between the (STAT5a+, ER α +) breast cancer subgroup and the menopausal status. The same was found for the different subgroups. In addition, the different subgroups are strongly associated with high tumor grades (SBR II and SBR III) but there is no significant association with node infiltration. The (STAT5a + ER α +) and (STAT5a + PR+) are associated with a proliferation index > 14% (P = 0.05 and P = 0.006, respectively). All subgroups are associated with a negative status of Her2.

The STAT5a mRNA level in (STAT5a + ER α +), (STAT5a + ER β +) and (STAT5a + PR+) breast cancer subgroups according to clinical and molecular information

Analysis of the STAT5a mRNA level, according to the clinical and molecular setting in the (STAT5a + ER α +) and (STAT5a + ER β +) breast cancer subgroups, shows the absence of any significant differences (Fig. 3a, b). Inversely, in the (STAT5a + PR+) subgroup, the STAT5a mRNA level increases for the postmenopausal status (P = 0.02) (Fig. 3c). In addition, a high

STAT5a mRNA level is associated with node infiltration (P = 0.03) (Fig. 3c).

Relationship between (STAT6 + ER α +), (STAT6 + ER β +) and (STAT6 + PR+) breast cancer subgroups and clinical settings

As shown in Table 4, there is the absence of a significant link between all subgroups and the menopausal status. Inversely, all subgroups are significantly associated with a high tumor grade. In addition, the coexpression of STAT6 with ER β (but not with ER α) is strongly associated with node infiltration (P = 0.001). Further, (STAT6 + PR+) is significantly associated with infiltration nodes (metastasis) (P = 0.03). A significant association of (STAT6 + ER α +) and (STAT6 + PR+) subgroups with a proliferation index > 14% (P = 0.03 and P = 0.006, respectively) and a strong negative association of all subgroups with HER2/status have been found.

The STAT6 mRNA level in (STAT5a + ER α +), (STAT5a + ER β +) and (STAT5a + PR+) breast cancer subgroups according to clinical and molecular information

When STAT6 is coexpressed with ER α , the increase of the STAT6 mRNA level is significantly associated with node infiltration (P = 0.02) (Fig. 3d). In the (STAT6 + ER β +) subgroup, the STAT6 mRNA level increases in the postmenopausal status (P = 0.01) (Fig. 3e). Finally, in the (STAT6 + PR+) breast cancer subgroup, increasing the STAT6 mRNA level was sig-

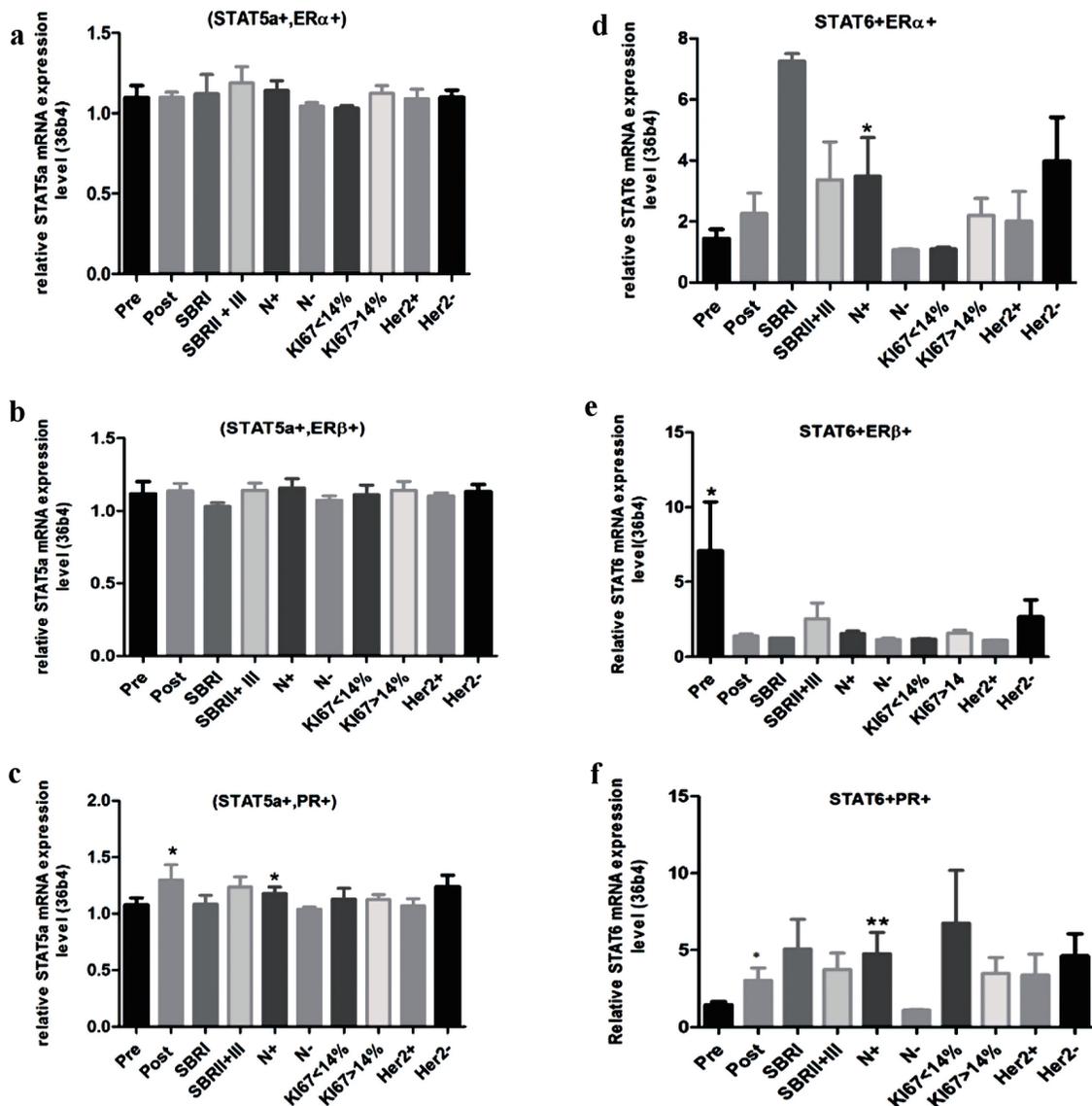


Figure 3. Relative mRNA expression level of STAT5a and STAT6 according to clinical and molecular information. (a) STAT5a mRNA expression level in (STAT5a+, ERα+) subgroup. (b) STAT5a mRNA expression level in (STAT5a+, ERβ+) subgroups. (c) STAT5a mRNA expression level in (STAT5a+, PR+) subgroups. (d) STAT6 mRNA expression level in (STAT6+, ERα+) subgroup. (e) STAT6 mRNA expression level in (STAT6+, ERβ+) subgroup. (f) STAT6 mRNA expression level in (STAT6+, PR+) subgroup. STAT: signal transducers and activators of transcription; ER: estrogen receptor; PR: progesterone receptor; SBR: Scarff-Bloom-Richardson.

nificantly associated with the postmenopausal status and with node infiltration (P = 0.05 and P= 0.001, respectively) (Fig. 3f).

Discussion

The present study shows that STAT5a and STAT6 are strongly expressed in malign breast tumors and can promote proliferation and metastasis in ERs and PR-positive breast cancer. In addition, positive STAT5a and STAT6 predict the response to endocrine therapy. We have shown here that STAT5a and

STAT6 were strongly overexpressed in malign breast tumors (87.5% and 78.12%, respectively) (Table 1). Our results agree with previous studies [8, 9]. There is no relationship between the STAT5a and STAT6 expressions and the menopausal status; however, considering the relative amount of mRNA, the STAT5a expression was found to be higher in the postmenopausal status compared to premenopausal breast tumors. Inversely, the mRNA expression level of STAT6 was much higher (three-fold) in premenopausal patients. Our finding shows a significant association of the STAT5a and STAT6 expressions with the histological tumor grade. This result is concordant with the findings of Yamashita et al who demonstrated that the

Table 4. Coexpression of STAT6 With ER α , ER β and PR and the Relationship With Clinical and Molecular Settings

	STAT6+					
	ER α + (n = 21)	P value	ER β + (n = 20)	P value	PR+ (n = 25)	P value
Menopausal status						
Pre-	9	NS	7	NS	12	NS
Post-	12		13		13	
SBR grade						
I	3	0.0000177	4	0.0005	6	0.0006
II + III	18		16		19	
Lymph node status						
N+	13	NS	15	0.004	17	0.02
N-	8		5		8	
KI67						
> 14	15	0.01	12	NS	18	0.004
≤ 14	6		8		7	
HER2						
+	16		16		6	
-	5	0.002	4	0.0005	19	0.0006

STAT: signal transducers and activators of transcription; ER: estrogen receptor; PR: progesterone receptor; SBR: Scarff-Bloom-Richardson; NS: not significant.

STAT5 expression was strongly correlated with the histological grade [19]. No links have been found between the STAT5a expression and lymph node metastasis. In this context, Wagner et al showed absence of a relationship between the STAT5a expression the menopausal status, the tumor size and metastasis [20]. On the contrary, the STAT6 expression is associated with node infiltration. This result is in agreement with a previous study that showed the association of STAT6 with node infiltration and distant metastasis [12, 21]. The analysis of the relative mRNA expressions of STAT5a and STAT6 shows the absence of a significant difference between node-positive and node-negative breast cancer. Inversely to previous data, our results reveal that STAT5a and STAT6 are both strongly associated with the tumor size. In addition, our results demonstrate a strongly negative association between the STAT5a and STAT6 expressions and the HER2/neu status. Consequently, STAT5 and STAT6 can predict the sensitivity to hormonal therapy. These results are in agreement with previous reports [19]. In addition, the present study reveals that there is no association between the STAT5a and STAT6 expressions and the proliferation index. This finding is concordant with the findings of Walker et al [22], which showed that the coactivation of STAT5 and STAT3 decrease cell proliferation in breast cancer, and with findings of Yu et al [23]. This showed that STAT5 negatively regulates cell proliferation by inducing cell cycle inhibitor genes.

Breast cancer is highly dependent on hormone receptors such as ER α , ER β and PR, which play a crucial role in the initiation and progression of the disease [14-16, 24]. The analysis of the STAT5a and STAT6 expressions compared to the hormonal receptor expressions revealed a significant positive association of STAT5a and STAT6 with ER α and PR. These

results are in agreement with previous studies which revealed an association of the STAT5 expression with the ER and PR positive statuses [19]. In addition, hyper-activation of STAT5a is associated with an increase of ER α [20]. The same is true for STAT6 which is expressed in ER+ breast cancer cell lines MCF-7 and ZR-75-1 [25].

The analysis of the STAT5a and the hormonal receptor (ER α , ER β and PR) coexpressions compared to the clinical and molecular settings shows no association with the menopausal status, whereas there are significant associations of all breast cancer subgroups with the tumor grade. In addition, the (STAT5a + ER α +) and (STAT5a + PR+) breast cancer subgroups are associated with a high proliferation index. Consequently, STAT5a can interact with ER α and PR to induce tumor progression and cancer cell proliferation. Our finding is in agreement with previous studies which showed that STAT5a may be activated by several signaling pathways such as the estrogen one [26]. In fact, activation of STAT5 by estrogen pathways through c-Src can promote cell proliferation and survival in breast cancer [26]. In fact, STAT5a regulates the gene expression that promotes cell proliferation and survival, such as a heat shock protein 90-A (HSP90A) and cyclin D1 [27]. The expression and activities of STAT5 are also upregulated by PR [28]. In addition, the PR - STAT5 interaction can regulate gene and protein expressions [29]. Furthermore, the coexpression of STAT5a with ERs and PR is associated with a negative status of Her2/neu. In fact, the present study shows a strongly negative correlation between the (STAT5a+; ER β +) and (STAT5a + PR+) breast cancer subgroups and the HER2/neu expression. A previous study showed links between Her2 downregulation and the sensitivity of endocrine therapy [30]. Consequently, the coexpression of STAT5a with ERs (especially ER β) and

PRs may be an indicator of endocrine sensitivity.

The same as it occurs with STAT5a, the coexpression of STAT6 with hormonal receptors (ER α , ER β and PR) is not associated with the menopausal status, while all breast cancer subgroups ((ER α +, STAT6+) (ER β +, STAT6+) (PR+, STAT6+)) are significantly associated with a high tumor grade (SBRII+ SBRIII). Moreover, the results of the present study demonstrate that an increase of the STAT6 mRNA expression is associated with infiltration nodes in (STAT6+, ER α +) and (STAT6+, PR+) breast cancer subgroups (Fig. 3b). In addition, the coexpression of STAT6 with ER β and PR is strongly associated with node infiltration (Table 4). Our findings are concordant with previous data that showed that STAT6 can affect tumor growth and metastatic niche formation [4-12, 21]. Furthermore, the association of the (STAT6+, ER β +) and (STAT6+, PR+) breast cancer subgroups with infiltration nodes and the (STAT6+, ER α +) and (STAT6+, PR+) breast cancer subgroups with a high proliferation index shows the possibility of crosstalk between STAT6 and ERs and PR. Finally, the results show a strongly negative association of all breast cancer subgroups with the HER2/neu expression. This may be an indication of the sensitivity to hormonal therapy. Indeed, such as with STAT5a, the coexpression of STAT6 with ERs and PR may be a good indicator for endocrine therapy in association with the downregulation of HER2.

In conclusion, the results of the present study demonstrate that the STAT5a and STAT6 expressions were retained in the majority of the breast cancer cases. Often, STAT5a and STAT6 were coexpressed with ERs (ER α and ER β) and PR. The coexpression of STAT5a and STAT6 with ERs and PR is associated with a high tumor grade, metastasis, and a high proliferation index. In fact, the (STAT5a+ ER α), (STAT5a + PR), (STAT6 + ER α) and (STAT6 + PR) breast cancer subgroups were associated with a high proliferation index while the (STAT6+ER β) and (STAT6+PR) breast cancer subgroups were associated with metastasis. This finding means that STAT5a and STAT6 can interact with ER α , ER β and PR and lead to cell proliferation and metastasis in malignant breast tumors.

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Conflict of Interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

Informed Consent

All the informed consents for publication were obtained.

Author Contributions

Mohamed Oueslati: study design, experimentation (immunohistochemistry and q-RT-PCR), results analysis and interpretation, writing of the manuscript. Ilhem Bettaieb determined the presence of malignant cells in the samples of breast cancer and did the analysis of immunohistochemistry results. Ridha Ben Younes contributed to the writing of the manuscript. Amor Gamoudi and Khaled Rahal determined the presence of malignant cells in the samples of breast cancer. Ridha Oueslati contributed to the results interpretation. All authors provided critical feedback and helped shape the research, analysis and manuscript.

Data Availability

The authors declare that data supporting the findings of this study are available within the article.

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